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Fractal approaches to characterize the structure of capillary suspensions using rheology and confocal microscopy

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Abstract

The rheological properties of a particle suspension can be substantially altered by adding a small amount of a secondary fluid that is immiscible with the bulk phase. The drastic change in the strength of these capillary suspensions arises due to the capillary forces, induced by the added liquid, leading to a percolating particle network. Using rheological scaling models, fractal dimensions are deduced from the yield stress and from oscillatory strain amplitude sweep data as function of the solid volume fraction. Exponents obtained using aluminum-oxide-based capillary suspensions, with a preferentially wetting secondary fluid, indicate an increase in the particle gel's fractal dimension with increasing particle size. This may be explained by a corresponding relative reduction in the capillary force compared to other forces. Confocal images using a glass model system show the microstructure to consist of compact particle flocs interconnected by a sparse backbone. Thus, using the rheological models two different fractal dimensionalities are distinguished – a lower network backbone dimension ($D = 1.86\text{--}2.05$) and an intrafloc dimension ($D = 2.57\text{--}2.74$). The latter is higher due to the higher local solid volume fraction inside of the flocs compared to the sparse backbone. Both of these dimensions are compared with values obtained by analysis of spatial particle positions from 3D confocal microscopy images, where dimensions between 2.43 and 2.63 are computed, lying between the two dimension ranges obtained from rheology. The fractal dimensions determined via this method corroborate the increase in structural compactness with increasing particle size.

Keywords

Capillary suspensions; Fractal dimension; Rheology; Confocal microscopy; Microstructure; Particle gels

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1 Introduction

Capillary suspensions are ternary solid-liquid-liquid systems. They consist of particle suspensions with a small amount of added secondary fluid that is immiscible with the bulk liquid [1]. The added fluid causes a sample-spanning particle network to form, which leads to a dramatic alteration of the suspension rheology and stability [1–6]. This effect can be used as a simple method to tune the flow behavior of suspensions that would otherwise require the addition of rheology control agents, surfactants or other additives [7]. Capillary suspensions can be used for various applications such as precursors for ceramic or glass filters with high porosity and simultaneously small pore size [8–10], porous polymeric or hybrid membranes [11, 12] improved polymer blends [13–15], slurries for battery electrodes or printable electronics with accurately molded edge shape, good charge transfer properties and reduced resistivity [16–18], crack-free films [19], thermal interface materials [20], carbon slurries from renewable sources used for energy generation [21], the preparation of novel food products [22, 23] or as 3D printing paste formulations [24, 25].

Capillary suspensions can be formed either with the secondary fluid wetting better than the bulk phase, which is referred to as pendular state, or wetting less well in the capillary state. In both cases, a percolating particle network arises leading to a transition in rheological properties due to network formation [1, 26]. Correspondingly, capillary suspensions can be regarded as attractive particle gels.

The structure of capillary suspensions and their strength depends on the solid particle radius a , the interfacial tension Γ between the fluids, and the wetting properties of both liquids comprised by the three phase contact angle θ that the secondary fluid forms towards the particles in a bulk liquid environment [7]. Additionally, structure and strength are determined by the volume and number of the second fluid entities interconnecting the particles [26]. The overall second fluid volume fraction ϕ_{sec} in capillary suspensions is small compared to the volume fraction of dispersed solid particles ϕ_{solid} and especially to the fraction of bulk liquid [6]. In the pendular state, the capillary force F_c in a concave bridge of volume V_{bridge} between two equally sized spherical particles can be computed either by solving the Young-Laplace equation or by assuming a certain bridge shape. Typically, the attraction energy between such particles connected by the capillary force is many times higher than thermal energy $k_B T$ [27]. Accordingly, the capillary force is also orders of magnitude higher than the van der Waals force [2] as well as gravity [28]. This leads to an increased cohesion between particles in capillary suspensions compared to suspensions without added secondary fluid that results in the observed marked changes in flow behavior and prevents particle sedimentation [22]. The capillary force of single bridges can be related to the macroscopic stress of a percolated capillary suspension network. The relationship between capillary force and yield stress σ_y for equally-sized particles in direct contact is given by

$$\sigma_y = f(\phi_{\text{solid}}, \phi_{\text{sec}}) \frac{F_c}{a^2} = f(\phi_{\text{solid}}, \phi_{\text{sec}}) g \left(\frac{V_{\text{bridge}}}{a^3} \right) \frac{2\pi\Gamma\cos\theta}{a} \quad (1)$$

with $f(\phi_{\text{solid}}, \phi_{\text{sec}})$ being a function of particle and second fluid volume fractions and $g(V_{\text{bridge}}/a^3)$ a function of the bridge volume relative to particle volume [2, 29–31]. While V_{bridge} is obviously related to the second fluid volume fraction ϕ_{sec} , it also strongly depends on the sample preparation method [13, 32] and contact angle [26]. Such a dependence on sample preparation conditions is true of capillary suspensions and other strong attractive particle gels that are not in thermodynamic equilibrium since the structure does not organize itself in a reproducible manner without the application of well-defined external forces [33].

The structure of attractive particle gels often is treated as a random fractal and thus is described by the fractal dimension D . The aim of this study is to apply fractal approaches to capillary suspensions and the paper is organized as follows. In section 2, we discuss the concept of fractal dimensions and summarize different approaches to calculate the fractal dimension of particle networks from rheological scaling laws or image analysis. After having described our materials and methods in section 3, our first study (section 4.1) evaluates the yield stress of aluminum oxide based capillary suspensions as a function of particle size as well as secondary and solid volume fractions. We then apply rheological scaling laws to evaluate fractal dimensions in section 4.2. In section 4.3, we deduce fractal dimensions from image analysis of the structure using a series of confocal microscopy experiments with capillary suspensions based on fluorescently labelled silica beads. Finally, we compare the approaches and summarize our findings in the conclusions of section 5.

2 Theoretical background

The fractal dimension D is a parameter to describe the internal geometry of particle clusters. When fractal clusters consisting of N particles are formed by aggregation, the cluster radius R_c scales as

$$N \propto R_c^D \quad (2)$$

where D in three-dimensional space can take values of $1 \leq D \leq 3$ [34, 35]. Counting the number of particles inside a sphere of radius R and transforming eq. 2 to non-dimensional form by relating the radial coordinate R to single particle radius a directly leads to

$$N(R/a) \propto (R/a)^D \quad (3)$$

where $N(R/a)$ is the particle number as function of the normalized radial coordinate R/a and the center of gyration of the cluster is defined as $R = 0$ [36]. In a real system, there is some variability in the size of the particles, the size of the aggregate, and the fractal dimension for each cluster, so these values should be taken as the mean values for the observed sample.

Different models have classically been used to describe cluster formation kinetics. The diffusion limited (DLCA) and reaction limited cluster aggregation (RLCA) models are distinguished by the sticking probability of each additional particle contacting the cluster. The sticking probability is 1 (every particle sticks to the cluster upon first contact) for

DLCA, while it is close to 0 for RLCA (a particle can contact the cluster many times before it finally adheres) [37]. Thus, RLCA clusters have a more densely packed structure than DLCA which is captured by their fractal dimension values. Typically, DLCA clusters have $D \approx 1.85 \pm 0.1$, while $D \approx 2.1 \pm 0.1$ for RLCA in three-dimensional space [37, 38]. However, this dimension can depend on a variety of other parameters, such as external shear [39, 40].

Experimental determination of D by eq. 3 is quite straightforward using 3D confocal images of the clustered particles from which the particle positions can be calculated [26, 41]. There are some caveats, however, when a percolated particle network instead of a single particle cluster is evaluated. In this case, different fractal clusters touch and merge into each other. Thus, adjacent clusters that were initially grown separately share particles with each other and the corresponding structure in the overlap regions can no longer be well described by the cluster fractal dimension. The correlation length ξ , which denotes the maximum length scale before a cluster loses its internal fractality, is a helpful tool to capture this critical overlap. Therefore, eqs. 2 and 3 are limited to $R < \xi$. Fig. 1 shows an idealized drawing of a particle gel originating from merged clusters where ξ is labeled. The correlation length decreases as the number density of particles is increased. For attractive particle gels, ξ can be directly related to the volume fraction of particles ϕ_{solid} , defined as the total particle volume divided by the total sample volume, with

$$\frac{\xi}{a} = \phi_{\text{solid}}^{1/(D-3)} \quad (4)$$

which also includes changes to the correlation length with the cluster structure via the dependence on D [42, 43]. While the cluster fractal dimension D is often treated as identical to the overall percolated gel fractal dimension, such fractality only holds for length scales smaller than ξ and the gel appears non-fractal and homogeneous at larger length scales [44]. Using the aforementioned approaches on larger length scales identifies an uncorrelated structure, resulting in $D = 3$ [43, 44].

Since the concept of fractality was introduced to characterize clustered structures of particles about three decades ago [45], a plethora of scaling theories have emerged that relate the fractal dimension to the rheological properties of attractive particle gels as a function of the particle volume fraction ϕ_{solid} (e.g. [36, 44, 46–53]). All of these scalings are of a power law type, such that

$$X(\phi_{\text{solid}}) \propto \phi_{\text{solid}}^{f(D)} \quad (5)$$

where X serves here as a placeholder for a rheological property, e.g. yield stress σ_y or elastic modulus plateau G_0 , and the scaling theories mostly differ in the exponent $f(D)$, commonly denoted by m . Piau et al [47] have proposed a model for the yield stress scaling of particle gels with

$$f(D) \equiv m = \frac{4}{3-D} \quad (6).$$

The model of Piau et al relates the yielding behavior of the particle gel to the free energy stored in a volume with scale ξ , which is not related to ϕ_{solid} via eq. 4, but instead via a scaling argument following the method of de Gennes designed for mesh size determinations of semi-dilute polymer gels [54]. While this derivation seems to be somewhat arbitrary, Piau et al have compared fractal dimensions calculated with their model to fractal dimensions experimentally determined by light scattering methods and found very good agreement. The Piau model has also been successfully applied to particle gels with a broad diversity of material compositions [47, 55–57].

Shih et al [46] proposed a very different model relating the network structure to the elastic properties of the particle gel, which are represented by two different elastic constants K_f and K_b . K_f is the intrafloc constant characterizing the elasticity inside a cluster of size ξ and K_b characterizes the interfloc or backbone connections between different flocs. The interfloc connecting regions are highlighted by the grey boxes in Fig. 1. Such an inhomogeneous gel structure is specifically expected for intermediate particle volume fraction ranges ($\phi_{\text{solid}} \approx 0.2\text{--}0.5$) [58, 59], which is often the relevant range for capillary suspensions. Following Shih et al, the relative contribution of both elastic constants determines the sign and slope of a scaling exponent $f(D) \equiv m_\gamma$ where the rheological property X in eq. 5 is the critical strain amplitude γ_{crit} from oscillatory shear rheometry that denotes the end of the linear viscoelastic deformation regime at a given frequency. A second scaling proposed for G_0 , the frequency-independent plateau value of the elastic modulus G' in the linear viscoelastic regime, has a scaling exponent $f(D) \equiv m_G$. The model of Shih et al has been further evaluated and extended by Wu and Morbidelli [48], who derived the two scaling exponents as

$$m_\gamma = \frac{2-\beta}{3-D} \quad (7)$$

$$m_G = \frac{\beta}{3-D} \quad (8)$$

with the parameter β containing information about the relative contributions of intrafloc and backbone elasticities. Eliminating β by combining eqs. 7 and 8 leads to an expression of D as a function of both scaling exponents

$$D = \frac{3(m_\gamma + m_G) - 2}{m_\gamma + m_G} \quad (9)$$

where D , following the definitions of eqs. 2 to 4, is the internal fractal dimension of the flocs.

The model of Piau et al, on the other hand, does not differentiate between the flocs and backbone as this theory originates from semi-dilute polymer network mesh sizes (i.e. small ϕ_{solid} for particle gels). Ergo, this model assumes the presence of a homogeneous backbone that resembles the structure of such polymer gels rather than densely packed flocs. Therefore, the corresponding fractal dimension calculated from Piau's model is not expected to be the same D as calculated from the other rheological models, but should rather correspond to a "backbone dimensionality" on larger length scales. This implies, then, a fractal network of clusters where the clusters of size ξ are the basic building blocks. Thus, ξ is the lower instead of the upper length scale for the fractal structure of the network backbone [60].

3 Materials and Methods

3.1 Materials

Commercial grade aluminum oxide particles ($\alpha\text{-Al}_2\text{O}_3$) were obtained from Almatix GmbH (Ludwigshafen, Germany) which were used to prepare samples for rheological characterization. Four types of Al_2O_3 particles with different average particle radius $a_{50,3}$ were used (CT3000SG, $a_{50,3} = 0.32 \mu\text{m}$; CT1200SG, $a_{50,3} = 0.70 \mu\text{m}$; CL3000SG, $a_{50,3} = 1.94 \mu\text{m}$, CT19FG, $a_{50,3} = 3.12 \mu\text{m}$). Fig. 2 shows the respective particle size distributions and SEM images of the particles. The particles have irregular, angular, convex shapes. Their density ρ is 3.94 g/mL. The bulk phase in the Al_2O_3 material system was paraffin oil (Carl Roth, Karlsruhe, Germany). It has a Newtonian flow behavior and a dynamic viscosity of $\eta(20^\circ\text{C}) = 0.029 \text{ Pa s}$. The density of the paraffin oil is 0.85 g/mL. As an appropriate secondary liquid phase, we used an aqueous sucrose solution (50 vol %) with $\rho(20^\circ\text{C}) = 1.28 \text{ g/mL}$ and $\eta(20^\circ\text{C}) = 0.080 \text{ Pa s}$. The D-(+)-sucrose was obtained from Carl Roth (Karlsruhe, Germany). The secondary liquid is no solvent for the $\alpha\text{-Al}_2\text{O}_3$. The interfacial tension between bulk and secondary liquids is $\Gamma = 41.2 \pm 0.7 \text{ mN/m}$ as determined with the pendant drop method (OCA 15 EC, Dataphysics, Filderstadt, Germany) and the contact angle of secondary fluid towards the Al_2O_3 in bulk environment is $\theta = 41 \pm 4^\circ$ as determined with the sessile drop method on a smooth $\alpha\text{-Al}_2\text{O}_3$ plate using the same apparatus as for pendant drop measurements. Thus, the system is in the pendular state.

For confocal microscopy, a different sample system based on silica particles was used. The silica particles were manufactured at Akzo Nobel (Bohus, Sweden; the smallest size donated by the manufacturer and the others purchased from MZ-Analysentechnik GmbH, Mainz, Germany). The silica particles were covalently dyed with fluorescent rhodamine B isothiocyanate via a modified Stöber synthesis [26, 61, 62] and the surface was partially hydrophobized with trimethylchlorosilane [63]. Our methods to perform these chemical particle reaction steps are detailed in the online supplementary information, section 1. Three types of silica with different mean particle radius were used (Kromasil 100-1.8-SIL, $a_{50,3} = 0.95 \mu\text{m}$; Kromasil 100-3.5-SIL, $a_{50,3} = 1.59 \mu\text{m}$; Kromasil 100-7-SIL, $a_{50,3} = 3.21 \mu\text{m}$; all sizes determined after chemical modifications). The dyed particles are spherical (see Fig. 2d) and the measured density is $1.95 \pm 0.08 \text{ g/mL}$.

The bulk phase in the SiO₂ material system was a mixture of 86.8 wt % 1,2-cyclohexane dicarboxylic acid diisononyl ester (Hexamoll DINCH, BASF, Ludwigshafen, Germany) and 13.2 wt % n-dodecane (Alfa Aesar, Karlsruhe, Germany). Bulk mixture properties are $\rho = 0.91$ g/mL and $\eta(20\text{ }^{\circ}\text{C}) = 0.021$ Pa s. The secondary phase in the SiO₂ material system was a mixture of 86.4 wt % glycerol (purity > 99.5 %, Carl Roth, Karlsruhe, Germany) and 13.6 wt % ultrapure water, which does not modify the silica particles. Minute amounts of the fluorescent dye PromoFluor-488 premium carboxylic acid (PromoKine, Heidelberg, Germany) were added to the secondary fluid. Secondary fluid mixture properties are $\rho = 1.23$ g/mL and $\eta(20\text{ }^{\circ}\text{C}) = 0.120$ Pa s. The bulk and secondary fluid mixtures had the same refractive index as the dyed silica particles ($n = 1.455 \pm 0.001$). Index matching was necessary to obtain high quality confocal images. The interfacial tension between bulk and secondary liquids, determined from pendant drop measurements, is $\Gamma = 21.5 \pm 0.3$ mN/m. Contact angles for all three silica particle sizes were directly determined from confocal images of individual particles that are situated in an interface between bulk and secondary liquid. A detailed description of this method is given elsewhere [26]. The contact angle of secondary fluid towards the dyed silica particles in bulk environment is $\theta = 65 \pm 7^{\circ}$. Thus, this system also is in the pendular state. This is a necessary precondition for assuring comparability of the silica and Al₂O₃ systems, together with the similar size ranges of both particle types. In the remainder of the paper, we will denote the mean particle radius $a_{50,3}$ simply by a .

3.2 Sample preparation

Suspensions based on Al₂O₃ particles were prepared by mixing the particles into the bulk phase with a high speed dissolver stirrer (diameter 5 cm) at a rotation speed of 1200 rpm for 10 min. Afterwards, the secondary liquid phase was added at a speed of 1000 rpm, and mixed for 2 min. After preparing the suspensions in the high speed dissolver, a homogenization step in a self-constructed ball mill (ball diameter 20-25 mm) followed. The homogenization time was 24 h at 18 rpm. The total sample volume was 100 mL.

The silica based capillary suspensions were prepared by emulsification of the secondary fluid in the bulk phase with an ultrasound sonifier (Ditigal Sonifier model 250, Branson Ultrasonics Corporation, Danbury, USA) with an ultrasonic horn of 12.5 mm diameter and maximum available output power of 200 W. Emulsification occurred at an amplitude of 35 % for 30 s. Thereafter, the particles were stirred in with a spatula and three more ultrasonic mixing steps were applied to the sample (each 10 s at 10 % amplitude, stirring with a spatula between steps to enhance sample homogeneity). The total sample volume was 0.85 mL. Preparation of large volumes was limited by the small total amounts of available silica particles resulting from the different chemical modification steps detailed in the online supplementary information, section 1. These small volumes are also the reason for not using the silica system for the extensive rheological measurements, where the Al₂O₃ system was chosen instead.

3.3 Rheological measurements

Rheological behavior of suspensions based on Al₂O₃ particles was determined with steady as well as oscillatory shear measurements. For yield stress determination, the stress-

controlled rheometers MARS II (minimum torque $M_{\min} = 50$ nNm) and RheoStress RS150 ($M_{\min} = 500$ nNm, both rheometers from Thermo Fisher Scientific, Karlsruhe, Germany) with a four blade vane geometry (5 mm radius, 22 mm blade height) in an appropriate cylinder cup (Z-20-DIN, 10.8 mm cup radius) were used and the shear stress was increased stepwise from 1 to 10,000 Pa. The steps were logarithmically distributed with 50 steps per decade. Each step followed a dwell time of 5 s. The capillary suspensions show transient behavior on short timescales, but the dwell time was monitored to be sufficient to reach steady state. The gels also do not demonstrate aging behavior over the experimental timescales. The yield stress was determined using the tangent method [64, 65]. The storage modulus G' and loss modulus G'' were determined via oscillatory shear measurements performed on the MARS II rheometer equipped with plate-plate geometry (diameter 35 mm, gap height 1 mm). Lower steel plate and upper titanium plate surfaces were sandblasted to prevent wall slip. Stress amplitude sweep measurements were performed at angular frequencies $\omega = 1$ rad/s and $\omega = 100$ rad/s. Stress controlled frequency sweep measurements were performed at a stress inside the linear viscoelastic region (sample dependent, between 10 Pa and 150 Pa) in a frequency range of 100-0.1 rad/s and show a frequency independent storage modulus of the suspensions in this range. All measurements were conducted at 20 °C. Despite the density mismatch between particles and liquids, neither particle sedimentation nor compaction of the sample during the rheological (and confocal) measurements is observed. This is due to the very strong capillary forces, which are orders of magnitude larger than the gravitational force on the particles.

3.4 Confocal microscopy

Confocal images were taken with a Leica TCS SP8 inverted confocal laser scanning microscope (Leica Microsystems, Mannheim, Germany). The microscope is equipped with two solid state lasers with wavelengths of 488 nm and 552 nm. PromoFluor 488 Premium carboxylic dye, in the capillary suspensions' secondary phase, is excited by the 488 nm laser and rhodamine B isothiocyanate, in the particles, is excited by the 552 nm laser. The secondary phase dye emission was detected in a wavelength range of 495-520 nm and the particles at 650-700 nm. A glycerol immersion objective with a numerical aperture of 1.3 and 63x magnification (HC PL APO 63x/1.30 GLYC CORR CS2, Leica Microsystems, Mannheim, Germany) was used for image acquisition.

Image analysis was completed using IDL (Exelis VIS). Localization of particle positions was based on a modification of the 2D particle tracking algorithm of Crocker and Grier [66], which had been further developed for 3D detection by Weeks [67, 68]. Due to the size distribution and high volume fractions of particles, some more elaborate processing steps were necessary, similar to the procedure used by Wenzl et al [69]. A detailed description of these procedures for particle localization and the subsequent determination of fractal dimensions from microscopy images is given in the online supplementary information, section 2.2.

4 Results and Discussion

4.1 Yield stress master curve for Al_2O_3 system

The yield stress σ_y of various samples of the Al_2O_3 system was measured with variations of the mean particle size a , secondary fluid volume fraction ϕ_{sec} and particle volume fraction ϕ_{solid} . Fig. 3a shows σ_y as function of ϕ_{sec} with different sizes a at a constant solid volume fraction $\phi_{\text{solid}} = 0.15$, while Fig. 3b shows σ_y as function of ϕ_{sec} with varied ϕ_{solid} at a constant particle size $a = 0.70 \mu\text{m}$. In all cases, the yield stress increases strongly at first with small amounts of added second fluid due to the buildup of capillary bridges in the pendular state, then goes through a maximum and finally σ_y decreases again when ϕ_{sec} is further increased. The position of the maximum yield stress is shifted to higher second fluid volumes with increasing solid volume fraction (Fig. 3b), but is independent of particle size (Fig. 3a). This maximum value is typically attributed to a funicular network structure, i.e. merged capillary bridges [26, 70], while the ensuing decrease occurs due to the formation of larger dense agglomerates, which lead to a weakening of the network structure [5, 9]. The details of yield stress dependence on second fluid content have already been thoroughly investigated by Dittmann et al on the same sample system [9], and are a general characteristics of pendular state capillary suspensions [71].

In Fig. 4, we show the same data as in Fig. 3 and include additional data taken at $\phi_{\text{solid}} = 0.10$ for $a = 0.32 \mu\text{m}$ as well as $\phi_{\text{solid}} = 0.20$ and 0.25 for $a = 3.12 \mu\text{m}$. This data has been rescaled so that all data collapse onto a single master curve that can be used to predict an approximate yield stress for any combination of particle size, second fluid and solid volume fraction. While this curve is a very helpful tool for directed product design based on such capillary suspensions, it is the details of the rescaling that are of fundamental scientific interest. The rescaling on the abscissa, where ϕ_{sec} is replaced by $\phi_{\text{sec}}/\phi_{\text{solid}}$, is quite easy to understand since all of the structural transitions, from pure suspension to pendular state to funicular state and finally to spherical agglomeration, depend on the relative amount of the secondary fluid droplets, characterized by droplet size and number, to the amount of particles. This ratio is captured by the relative volume fractions with the addition of the average number of bridges per particle. Increasing, e.g., the particle volume fraction while keeping the second fluid volume fraction unchanged would either, at fixed droplet volumes, lead to a reduced number density of pendular bridges per particle contacts or, retaining the bridge number density, to a reduced volume of the bridges, and thus a different network structure or mechanical strength. Indeed, a mixed contribution of both effects is likely to appear. Accordingly, $\phi_{\text{sec}}/\phi_{\text{solid}}$ has already been used as relevant property involved in structural transitions of capillary suspensions in other studies [5, 20, 65, 71]. The transition from the yield stress increase to the maximum plateau value at $\phi_{\text{sec}}/\phi_{\text{solid}} \approx 0.02\text{--}0.04$ agrees well with the ratio of single bridge to particle volumes $V_{\text{bridge}}/V_{\text{particle}} = 0.021\text{--}0.025$ where the pendular-to-funicular transition was calculated for glass bead based capillary suspensions with contact angles in the range of $\theta = 40\text{--}61^\circ$ [26]. Similarly, for wet granular materials with $\phi_{\text{solid}} \approx 0.57$ the upper limit of the pendular regime has been found at $\phi_{\text{sec}} \approx 0.025$, denoting this transition to take place at $\phi_{\text{sec}}/\phi_{\text{solid}} \approx 0.04$ [70].

The rescaling on the ordinate axis of Fig. 4 made by dividing the yield stress by $\phi_{\text{solid}}^{3.7}$ and $a^{-0.6}$ is more complex to interpret. Using eq. 1 as a guide, the dependence on V_{bridge} is via the normalized second fluid ratio $\phi_{\text{sec}}/\phi_{\text{solid}}$ on the abscissa and Γ and θ are constant for all samples. Therefore, σ_y can be rescaled by the only two remaining variables ϕ_{solid} and a . From eq. 1, σ_y should then be a function of a^{-1} instead of $a^{-0.6}$ and the solid volume scaling is comprised in the unknown term $f(\phi_{\text{solid}})$. Recently, $f(\phi_{\text{solid}}) = \phi_{\text{solid}}^2$ from sparse suspensions was proposed by Koos [2], but this obviously cannot hold. A dependence of $\phi_{\text{solid}}^{3.3}$ has been found by Domenech et al for a capillary suspension system with a much lower θ [5], which corroborates the dependence of $\phi_{\text{solid}}^{3.7}$ found here. Additionally, we have performed yield stress measurements with another capillary suspension system based on CaCO_3 particles, presented in the online supplementary information, section 3, that can also be rescaled using a solid volume fraction exponent of 3.7, which is further confirmation for the presented result.

While the theoretical size dependence of a^{-1} already has been verified for capillary suspensions using glass beads [7], the deviation here might be explained by the presence of particle aggregates. It may be assumed that these aggregates rather than the individual particles form a percolated aggregate network similar to what was proposed towards the end of section 2. These aggregates should then form a network backbone and the yield stress is related to this network backbone rupturing. The size of the aggregates varies based on the relative droplet volume, so the exponent cannot be easily predicted and the value of -0.6 here was empirically determined. An example estimation of exponents closer to -1 using agglomerate sizes is shown in the online supplementary information, section 4. However, other explanations for the deviation from the a^{-1} scaling are possible, e.g. a different polydispersity in particle size distributions or deviations from ideal spherical particle shape as seen in Fig. 2a and b.

4.2 Fractal dimension from rheological scaling laws

For capillary suspension based product design, the maximum achievable yield stress value is often of salient interest. As can be seen in Fig. 4, this maximum appears in the Al_2O_3 system around a fixed sample composition ratio of $\phi_{\text{sec}}/\phi_{\text{solid}} = 0.12$. Fig. 5 shows the yield stress at this fixed ratio as a function of the solid volume fraction for all four particle sizes. While each particle size clearly shows a power law scaling, a closer look reveals that the scaling exponent of 3.7 used for the master curve is only an inaccurate averaged value. The actual exponents m increase with increasing particle sizes from $m = 3.50 \pm 0.18$ at $a = 0.32 \mu\text{m}$ to $m = 4.23 \pm 0.13$ at $a = 3.12 \mu\text{m}$.

In Fig. 6, the Piau model (eq. 6) is applied to these scaling exponents to calculate the fractal dimensions D as function of the particle size. The model predicts fractal dimensions increasing from $D = 1.86 \pm 0.06$ at $a = 0.32 \mu\text{m}$ to $D = 2.05 \pm 0.03$ at $a = 3.12 \mu\text{m}$. These values all lie between the values of 1.85 and 2.10 reported for the DLCA and RLCA aggregation kinetic models [38]. However, one should hesitate when directly applying the DLCA and RLCA models from purely Brownian particle motion to capillary suspensions that are prepared using strong mixing (i.e. external flow fields determine particle motion) and are also influenced by gravity. In literature, hydrodynamic forces [72, 73] and gravity

[74–76] both have been shown to lead to rearrangements of clusters with more compact geometries with higher D compared to purely diffusive aggregates, and a loss of fractality in some cases. This would imply that we should observe higher fractal dimensions $D > 2.1$ that do not lie in the range between DLCA and RLCA values. As mentioned in the end of the theory section 2, however, the Piau model is expected to predict the dimension of a network's backbone instead of the internal dimension of the basic aggregates, which may explain the calculated fractal dimensions in Fig. 6 in this range. External forces mainly influence the internal structure of the aggregates and should have less influence on the percolating backbone as this structure only obtains its final configuration once the rapid external mixing has ceased. This backbone by itself is relatively sparse compared to the interior of the aggregate, even if there is an interconnected structure. Thus, the backbone dimensionality shown in Fig. 6 should be lower than the internal aggregate dimensionality, as fractal dimensions are linked to the relative number density of particles (eq. 3) [43, 77, 78].

By considering the dependence of our observed dimensionalities on particle size in terms of aggregation, the sticking probability for approaching particles decreases with larger particle sizes. A hypothesis to explain this finding can be drawn when the attractive capillary force F_c is compared to other forces acting on the colliding particles. While F_c is a linear function of particle size $F_c \sim a$ (eq. 1), the inertial force F_i and hydrodynamic force F_h scale as $F_i \sim a^3$ and $F_h \sim a^2$, respectively [28]. Thus, with increasing particle size, the ratio of the capillary to inertial and to hydrodynamic forces becomes weaker, which leads to a reduced sticking probability and thus to more compact agglomerate structures. The ratio of capillary to the other relevant forces decreases with increasing particle size independent of the mode of aggregation. Therefore, the increase in the fractal dimension for capillary suspensions with increasing particle size, seen in Fig. 6, is also expected even in the present case where the aggregation is not governed by diffusion. Further corroboration for this increase is given by yield stress data obtained with another sample system based on CaCO_3 particles, which is shown in the online supplementary information, section 3.

It is also possible to determine the fractal dimension applying the model of Wu and Morbidelli to data from oscillatory shear rheometry. Fig. 7 shows an example measurement of the moduli as a function of frequency as well as strain amplitude for an Al_2O_3 -based capillary suspension at $\phi_{\text{sec}}/\phi_{\text{solid}} = 0.12$. All samples, independent of ϕ_{solid} and a , showed a frequency independent elastic modulus G' at strain amplitudes within the linear viscoelastic region for the frequency range tested (Fig. 7a). Therefore, the frequency independent plateau modulus G_0 can be identified directly using an amplitude sweep as shown in Fig. 7b. In addition to G_0 , a critical strain amplitude γ_{crit} denoting the limit of linear viscoelastic response was determined following Shih et al where γ_{crit} is defined using the 95 % criterion [46]. The critical amplitude showed no dependence on frequency ω at which the amplitude sweep was measured. The plateau modulus G_0 and the amplitude γ_{crit} are shown in Fig. 8a and Fig. 8b, respectively, as function of ϕ_{solid} for the four different particle sizes. G_0 can, with good agreement, be fitted by a power law with exponent m_G in the range of 3.46 ± 0.26 to 4.27 ± 0.34 , but no trend with particle size is observed. The γ_{crit} data is more scattered, but still is fit using simple power law, as often done in literature [46, 48, 79]. The exponents

increase with increasing particle size from $m_\gamma = 1.11 \pm 0.20$ at $a = 0.32 \mu\text{m}$ to $m_\gamma = 4.28 \pm 1.05$ at $a = 3.12 \mu\text{m}$.

The fractal dimensions D calculated from m_G and m_γ using the model of Wu and Morbidelli [48] (eq. 9) are shown in Fig. 8c for all four particle sizes. In contrast to the fractal dimensions calculated with the Piau model from the yield stress scalings (Fig. 6), these dimensions are expected to represent the internal floc structure and not the network backbone. The values are significantly higher than the RLCA value of $D = 2.1$, which due to the elevated solid volume fraction and strong external forces during sample preparation is expected. Although these dimensions differ from the ones calculated from the yield stress (Fig. 6), the trend of an increasing D with particle size is reproduced. Here, the dimension ranges from $D = 2.57 \pm 0.03$ at $a = 0.32 \mu\text{m}$ to $D = 2.74 \pm 0.04$ at $a = 3.12 \mu\text{m}$.

Finally, it is worth noting some peculiarities in the scaling of G_0 and γ_{crit} with particle size. First, the dependence of the plateau modulus scaling exponents, seen in Fig. 8a, is not directly expected. While G_0 is known to often be independent of particle size for a uniform gel structure [80] this is not obvious for the heterogeneous gels investigated here. The internal floc packing geometry should depend on the particle size, especially when the agglomerating tendencies of a single particle, which was discussed at the end of section 4.1 and is shown in Fig. S5 of the supplementary information, are kept in mind. Thus, since the fractal dimension is expected to increase, we might expect the G_0 scaling exponents to also increase with particle size. This trend is clearly shown in the scaling exponents for the critical strain (Fig. 8b) and for the yield stress (Fig. 5). The model from Wu and Morbidelli can help to clarify this issue. The values of m_G and m_γ , obtained from Fig. 8a and 8b, can be used to estimate not only D , but also β using eq. 7 and eq. 8. This parameter β denotes the relative strength of inter- and intrafloc connections in the network and decreases from $\beta \approx 1.5 \pm 0.2$ at $a = 0.32 \mu\text{m}$ to $\beta \approx 0.9 \pm 0.2$ at $a = 3.12 \mu\text{m}$. A diagram of β as function of the Al_2O_3 particle size is provided in Fig. S6 of the supplementary information. The limiting cases for β are the pure weak-link behavior (interfloc links are much weaker than the intrafloc links), corresponding to $\beta = 1$, and strong-link behavior (interfloc links are stronger than the intrafloc ones), where $\beta \approx 4.3$. These limits were first proposed in the original model of Shih et al [46] with the intermediate values assigned only later by Wu and Morbidelli [48]. Therefore, the comparably low β values in our study show that our samples are primarily dominated by weak interfloc links in all cases, but that these still become relatively weaker with increasing particle size, reaching the limit of pure weak-link behavior. The local change in packing inside of the flocs with increasing particle size (i.e. the increase in D) does not correspond to an increase in the plateau modulus since the strengthening inside of the flocs is counteracted by a weakening of the interfloc bonds. The critical strain amplitude, however, is more sensitive to the weakening of interfloc bonds, which causes the observed dependence of the corresponding scaling exponents m_γ on particle size, as seen in Fig. 8b.

4.3 Structural investigations using confocal microscopy

We can image capillary suspensions using a model system to see the differences between the compact clusters and sparse backbone. 2D confocal images of capillary suspensions based

on silica particles with three different sizes a and volume fraction $\phi_{\text{solid}} = 0.26 \pm 0.06$ are shown in Fig. 9. Following the results for the Al_2O_3 system, the ratio $\phi_{\text{sec}}/\phi_{\text{solid}} = 0.12$ was also kept constant for all confocal samples. The three confocal images have been rescaled in size to allow better visual comparability so that the scale bar in each of the three images denotes a length of $40 \cdot a$. These images do not show particle gels that are homogeneous at these scales, but a structure consisting of relatively compact agglomerates with large (sparsely populated) voids. Thus, the model structure shown in Fig. 1 is indeed an appropriate representation of the structure of capillary suspensions. This means that making a distinction in the strength and structure between the backbone and agglomerate obtained using the two rheological models in section 4.2 is appropriate.

The total number of particles N around a reference particle in the 3D images can be determined as function of the normalized radial distance R/a to the reference particle by detecting the particle positions. An example 3D image and the method for determining the positions is described in section 2 in the online supplementary information. This function $N(R/a)$ can be used to calculate the fractal dimension using eq. 3. Since the clusters have a limited size, $N(R/a)$ is not calculated for distances longer than the correlation length ξ/a , which is estimated from eq. 4 using the dimension D from the rheological results at first. The lowest floc dimension calculated from the oscillatory data using the Wu and Morbidelli model in section 4.2 was $D \approx 2.6$. The measured $\phi_{\text{solid}} = 0.26 \pm 0.06$ leads to $\xi/a \approx 29 \pm 12$ (with the error in ξ/a being estimated based on the error in ϕ_{solid} and not in D). This value roughly matches the cluster sizes visible in Fig. 9. When using the same ϕ_{solid} , but $D = 2.0$, which is in the range predicted from the Piau model, the calculated correlation length drastically shrinks to $\xi/a \approx 3.85 \pm 0.65$. However, using $\phi_{\text{solid}} = 0.26$ is incorrect in the latter calculation. While the relevant solid volume fraction for the backbone cannot be directly defined, the number of contributing particles is clearly lower than the total particle number in the sample. Thus, the relevant ξ/a value for the backbone should be larger in reality.

$N(R/a)$ functions have been determined from 3D confocal images with image sizes of $(45 \cdot a)^3$. These images contain an average of 3300 ± 850 particles and the corresponding local particle volume fraction $\phi_{\text{solid}} = 0.26 \pm 0.06$ was directly evaluated from the 3D images. Images used for fractal dimension determination have been taken at a much higher resolution than shown in Fig. 9, and the secondary fluid was also imaged. An example image is provided in the online supplementary information, section 2.1. Due to the large variation in volume fraction and particle number caused by the network inhomogeneity, as seen in Fig. 9, the $N(R/a)$ function is strongly influenced by the position inside of the sample where these relatively small 3D images are taken. Furthermore, the arbitrarily chosen reference particles ($R = 0$) also cannot be expected to coincide with the center of gyration of a cluster. Therefore, eight such images were taken for each particle size and the $N(R/a)$ function in each image was calculated using between 350 and 550 reference particles, randomly distributed in the volume not intersecting the edges, to decrease the influence of these errors. Additionally, the maximum radial distance was set to $R/a = 10$, considerably lower than the estimated cluster size of $\xi/a \approx 29 \pm 12$, in order to ensure many of these calculations were made inside of a cluster. The mean $N(R/a)$ functions of all these calculations are shown in Fig. 10 for the three different particle sizes.

The mean $N(R/a)$ functions show a power law increase where the exponent is evaluated for $2 < R/a < 10$ and the fractal dimension D equals the exponent following eq. 3. The dimensions, calculated from the image sets for each particle size, are shown in Fig. 11 as red diamonds.

This method captures mixed contributions from both the internal aggregate as well as the backbone structure as not all reference particles are close to the gyration center of the clusters, and can also include a portion of the voids, thereby reducing the effective particle density. Thus the optically determined D values are larger than the calculations from the rheological scaling model of Piau, but smaller than the ones from the Wu and Morbidelli scaling model, representing backbone and aggregate dimensions, respectively. The inhomogeneity of the particle networks leads to scattered data in the calculations of D from $N(R/a)$ functions, as local structures will vary significantly between different 3D images and around different reference particles. This scatter is seen in the significant size of the error bars in Fig. 10 and 11. As with the dimension determined from rheological data, the fractal dimension values from microscopy image analysis increase with increasing particle size. This is true even though a different method and sample system were used here. There is an increase from $D = 2.43 \pm 0.17$ at $a = 0.95 \mu\text{m}$ to $D = 2.63 \pm 0.07$ at $a = 3.21 \mu\text{m}$. Corresponding correlation lengths can be estimated, lying between $11 \lesssim \xi/a \lesssim 38$.

Some caveats should be kept in mind when directly comparing the three curves of Fig. 11, as there are some differences between the data obtainment from rheology and from confocal microscopy. Most obvious is the usage of two sample systems, especially the difference between perfectly spherical glass beads and more irregularly shaped Al_2O_3 particles. Also, different mixing protocols had to be used to prepare the samples, which has been recently shown to influence the strength of capillary suspensions [13, 32]. Closely related to the issue of mixing protocols is the general pre-shear history of the samples, including the loading into the measuring device for the rheometer which also differs from the microscope slide. Such different pre-shear protocols may also lead to different structures. Indeed, the rheological response itself might be altered by this pre-shear, as was shown for shear rate dependent thixotropic time scales in silica suspensions by Dullaert and Mewis [81]. Thus, comparing the absolute values of the fractal dimensions determined through confocal microscopy with the values deduced from rheology may not be valid. However, the approximate values and trends in the fractal dimensions from the different approaches are still expected to be valid. It is clear from both the rheological data and the microscopic data that these capillary suspensions exhibit a heterogeneous gel structure with fractal dimensions that increase with increasing particle size.

5 Conclusions

In this study, we used several approaches to determine the fractal dimension of capillary suspensions. We used rheological scaling laws that based on different structural and aggregation models and analyzed yield stress data or the elastic plateau modulus and the critical strain amplitude. These two different approaches led to different fractal dimensions capturing structural features on different length scales of the particle networks. Fractal dimensions in the range from 1.86 to 2.05 were determined using the rheological scaling

model from Piau et al [47] assuming a uniform fractal structure, while distinctly higher D values in the range from 2.57 to 2.74 were obtained for the same capillary suspensions from the rheological scaling model of Wu and Morbidelli [48]. This marked difference is explained by an inhomogeneous microstructure of capillary suspensions, where relatively dense agglomerates are linked to form a relatively sparse backbone transmitting the forces. Taking into account the origin of the two applied rheological scaling models, the fractal dimension calculated with the Piau model is then expected to be a measure of the network backbone structure while the model from Wu and Morbidelli describes the fractal dimension inside the aggregates. Due to the locally higher particle volume fraction and more compact structure, the floc dimension is much higher than the backbone dimension.

This hypothesis for interconnected dense flocs is confirmed by confocal images where particle-laden areas are separated by voids with few particles. Fractal dimensions were directly determined from particle positions obtained from 3D confocal images of index-matched capillary suspension model systems. This approach is generally more straightforward since the structural parameter D is directly obtained rather than the indirect method of deducing D from rheological data. While such a direct approach avoids complications and questions regarding the approximations and assumptions made to relate structure to rheology, it does not necessarily capture the inhomogeneity of the network structure. Based on structural considerations of the inhomogeneous sample structure, we indeed think it is more appropriate to not describe capillary suspension structures with only one single fractal dimension value, but to make distinctions between two different dimensionalities – one for the backbone at long length scales, and one for the internal aggregate dimensionality at short length scales. This latter value should be the parameter most closely related to the classical definition of fractal dimensions (eq. 2). Due to the presence of such different length scales, the direct approach to determine a single dimension D using particle number functions from microscopy also can only provide an apparent fractal dimension containing mixed contributions of both length scales. Due to the short length $R/a < 10$ used to evaluate the data, the contribution of the internal cluster structure is expected to dominate, however. Accordingly, the optically determined D values lie between the results obtained from the different rheological approaches, but closer to the intrafloc values from the model of Wu and Morbidelli than to the backbone dimensions captured by the Piau model.

Regardless of the method used, the fractal dimension of capillary suspensions increases with increasing particle size. This trend is confirmed by all of the sample systems at the different length scale provided by each method. Therefore, capillary suspensions with larger particles exhibit more compact particle arrangements. This can be explained by the weakening of the capillary force in comparison to both the inertial and hydrodynamic forces with increasing particle size. This should reduce the sticking probability of an incoming particle attaching to an aggregate during the preparation of the capillary suspension samples.

The inhomogeneous sample structure and refinement of the optically determined fractal dimension should be investigated further. In particular, our study has shown that the fractal dimension might not be the most meaningful property to get a comprehensive and complete insight into the microstructure of these capillary suspensions due to their heterogeneous

nature. Other characterization approaches provide additional options that can be combined with the fractal dimension to get structural information for such heterogeneous, high volume fraction particle gels. One promising method is that of Wyss et al, who evaluated a similar heterogeneous particle gel by describing it as two-component system with one phase being represented by large voids of bulk liquid and the second component by the compact particle gel (aggregates and backbone) [59]. This second component is described by a solid volume fraction ϕ_2 that is larger than ϕ_{solid} and is calculated as $\phi_2 = \phi_{\text{solid}}/(1-\phi_1)$ with ϕ_1 representing the estimated volume fraction of the void pores of bulk liquid. While it is difficult to precisely determine ϕ_1 , it nevertheless should be possible to estimate this value from confocal images in future studies and then determine local aggregate structures represented by ϕ_2 . A second promising method to characterize heterogeneous suspensions, first introduced by Varadan and Solomon, is based on the determination of Voronoi volumes around the individual particles. The broadness and the skewness of the distribution of Voronoi volumes can be used to identify the degree of structural heterogeneity [82]. One can also use other parameters to describe the network structure, such as the coordination number between particles evaluated from the confocal images [83] and then make distinctions between the particle contacts with and without liquid bridges. Similarly, obtaining a distribution of clustering coefficients might be a worthwhile approach to the local microstructure. Concerning rheological methods, detections of space-resolved stress distributions could help in fathoming details of the yielding mechanism, as yielding of heterogeneous gels can occur by only very local breaking of a very small fraction of the network connections [84]. This might also help to better explain the relative importance and strength of inter- and intrafloc bonds, which has emerged in the discussion of Fig. 8. Finally, another promising approach could be to perform simulations of the network structure and compare these simulations to optically determined data. This approach would allow structural information to be determined as sample conditions are independently varied. Thus, while we have gained interesting information about the structure of capillary suspensions by using the fractal approaches in this study, there remain many additional options for promising future research studies.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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References

- [1]. Koos E, Willenbacher N. Capillary forces in suspension rheology. *Science*. 2011; 331:897–900. [PubMed: 21330542]
- [2]. Koos E. Capillary suspensions: Particle networks formed through the capillary force. *Current Opinion in Colloid & Interface Science*. 2014; 19:575–684. [PubMed: 25729316]

- [3]. van Kao S, Nielsen LE, Hill CT. Rheology of concentrated suspensions of spheres. II. Suspensions agglomerated by an immiscible second liquid. *Journal of Colloid and Interface Science*. 1975; 53:367–373.
- [4]. McCulfor J, Himes P, Anklam MR. The effects of capillary forces on the flow properties of glass particle suspensions in mineral oil. *AIChE Journal*. 2011; 57:2334–2340.
- [5]. Domenech T, Velankar SS. On the rheology of pendular gels and morphological developments in paste-like ternary systems based on capillary attraction. *Soft Matter*. 2015; 11:1500–1516. [PubMed: 25582822]
- [6]. Velankar S. A non-equilibrium state diagram for liquid/fluid/particle mixtures. *Soft Matter*. 2015; 11:9393–8403. [PubMed: 26439643]
- [7]. Koos E, Johannsmeier J, Schwebler L, Willenbacher N. Tuning suspension rheology using capillary forces. *Soft Matter*. 2012; 8:6620–6628.
- [8]. Dittmann J, Koos E, Willenbacher N. Ceramic capillary suspensions: Novel processing route for macroporous ceramic materials. *Journal of the American Ceramic Society*. 2013; 96:391–397.
- [9]. Dittmann J, Willenbacher N. Micro structural investigations and mechanical properties of macro porous ceramic materials from capillary suspensions. *Journal of the American Ceramic Society*. 2014; 97:3787–3792.
- [10]. Maurath J, Dittmann J, Schultz N, Willenbacher N. Fabrication of highly porous glass filters using capillary suspension processing. *Separation and Purification Technology*. 2015; 149:470–478.
- [11]. Dittmann J, Maurath J, Bitsch B, Willenbacher N. Highly Porous Materials with Unique Mechanical Properties from Smart Capillary Suspensions. *Advanced Materials*. 2016; 28:1689–1696. [PubMed: 26677099]
- [12]. Hauf K, Riazi K, Willenbacher N, Koos E. Radical polymerization of capillary bridges between micron-sized particles in liquid bulk phase as a low temperature route to produce porous solid materials. *Colloid & Polymer Science*. 2017; 295:1773–1785.
- [13]. Domenech T, Velankar S. Capillary-driven percolating networks in ternary blends of immiscible polymers and silica particles. *Rheologica Acta*. 2014; 53:593–605.
- [14]. Heidlebaugh S, Domenech T, Iasella S, Velankar SS. Aggregation and separation in ternary particle/oil/water systems with fully-wettable particles. *Langmuir*. 2014; 30:63–74. [PubMed: 24345163]
- [15]. Dapeng W, Wang X, Yuan Y, Li W, Tian H, Zhao S. Increasing the apparent shear viscosity of polymer composites by uptake of a small amount of water. *RSC Advances*. 2014; 4:24686–24691.
- [16]. Bitsch B, Dittmann J, Schmitt M, Scharfer P, Schabel W, Willenbacher N. A novel slurry concept for the fabrication of lithium-ion battery electrodes with beneficial properties. *Journal of Power Sources*. 2014; 265:81–90.
- [17]. Bitsch B, Gallasch T, Schroeder M, Börner M, Winter M, Willenbacher N. Capillary suspensions as beneficial formulation concept for high energy density Li-ion battery electrodes. *Journal of Power Sources*. 2016; 328:114–123.
- [18]. Schneider M, Koos E, Willenbacher N. Highly conductive, printable pastes from capillary suspensions. *Scientific reports*. 2016; 6 31367.
- [19]. Schneider M, Maurath J, Fischer S, Weiß M, Willenbacher N, Koos E. Suppressing crack formation in particulate systems by utilizing capillary forces. *ACS Applied Materials and Interfaces*. 2017; 9:11095–11105. [PubMed: 28263554]
- [20]. Sun H, Zhang X, Yuen MM. Enhanced conductivity induced by attractive capillary force in ternary conductive adhesive. *Composites Science and Technology*. 2016; 137:109–117.
- [21]. Jampolski L, Sängler A, Jakobs T, Guthausen G, Kolb T, Willenbacher N. Improving the processability of coke water slurries for entrained flow gasification. *Fuel*. 2016; 185:102–111.
- [22]. Hoffmann S, Koos E, Willenbacher N. Using capillary bridges to tune stability and flow behavior of food suspensions. *Food Hydrocolloids*. 2014; 40:44–52.
- [23]. Wollgarten S, Yuce C, Koos E, Willenbacher N. Tailoring flow behavior and texture of water based cocoa suspensions. *Food Hydrocolloids*. 2016; 52:167–174.

- [24]. Maurath J, Willenbacher N. 3D printing of open-porous cellular ceramics with high specific strength. *Journal of the European Ceramic Society*. 2017; 37:4833–4842.
- [25]. Roh S, Parekh DP, Bharti B, Stoyanov SD, Velev OD. 3D Printing by Multiphase Silicone/Water Capillary Inks. *Advanced Materials*. 2017; 29:1701554.
- [26]. Bossler F, Koos E. Structure of particle networks in capillary suspensions with wetting and non-wetting fluids. *Langmuir*. 2016; 32:1489–1501. [PubMed: 26807651]
- [27]. Mitarai N, Nori F. Wet granular materials. *Advances in Physics*. 2006; 55:1–45.
- [28]. Herminghaus, S. *Wet Granular Matter: A Truly Complex Fluid*. World Scientific; Singapore: 2013.
- [29]. Willett CD, Adams MJ, Johnson SA, Seville JP. Capillary bridges between two spherical bodies. *Langmuir*. 2000; 16:9396–9405.
- [30]. Strauch S, Herminghaus S. Wet granular matter: A truly complex fluid. *Soft Matter*. 2012; 8:8271–8280.
- [31]. Megias-Alguacil D, Gauckler LJ. Capillary forces between two solid spheres linked by a concave liquid bridge: Regions of existence and forces mapping. *AIChE Journal*. 2009; 55:1103–1109.
- [32]. Bossler F, Weyrauch L, Schmidt R, Koos E. Influence of mixing conditions on the rheological properties and structure of capillary suspensions. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2017; 518:85–97. [PubMed: 28194044]
- [33]. Butt H-J. Controlling the flow of suspensions. *Science*. 2011; 331:868–869. [PubMed: 21330524]
- [34]. Jullien R. Aggregation phenomena and fractal aggregates. *Contemporary Physics*. 1987; 28:477–493.
- [35]. Witten T Jr, Sander LM. Diffusion-limited aggregation, a kinetic critical phenomenon. *Physical review letters*. 1981; 47:1400–1403.
- [36]. De Rooij R, Potanin A, Van den Ende D, Mellema J. Steady shear viscosity of weakly aggregating polystyrene latex dispersions. *The Journal of chemical physics*. 1993; 99:9213–9223.
- [37]. Kolb M, Jullien R. Chemically limited versus diffusion limited aggregation. *Journal de Physique Lettres*. 1984; 45:977–981.
- [38]. Lin M, Lindsay H, Weitz D, Ball R, Klein R, Meakin P. Universality in colloid aggregation. *Nature*. 1989; 339:360–362.
- [39]. Sonntag RC, Russel WB. Structure and breakup of flocs subjected to fluid stresses: I. Shear experiments. *Journal of colloid and interface science*. 1986; 113:399–413.
- [40]. Krall A, Weitz D. Internal dynamics and elasticity of fractal colloidal gels. *Physical review letters*. 1998; 80:778.
- [41]. Dinsmore AD, Weeks ER, Prasad V, Levitt AC, Weitz DA. Three-dimensional confocal microscopy of colloids. *Applied Optics*. 2001; 40:4152–4159. [PubMed: 18360451]
- [42]. Mobuchon C, Carreau PJ, Heuzey M-C. Structural analysis of non-aqueous layered silicate suspensions subjected to shear flow. *Journal of Rheology*. 2009; 53:1025–1048.
- [43]. Mellema M, Van Opheusden J, Van Vliet T. Relating colloidal particle interactions to gel structure using Brownian dynamics simulations and the Fuchs stability ratio. *The Journal of chemical physics*. 1999; 111:6129–6135.
- [44]. Bremer LG, van Vliet T, Walstra P. Theoretical and experimental study of the fractal nature of the structure of casein gels. *Journal of the Chemical Society, Faraday Transactions 1: Physical Chemistry in Condensed Phases*. 1989; 85:3359–3372.
- [45]. Jullien, R., Botet, R. *Aggregation and fractal aggregates*. World Scientific; Singapore: 1987.
- [46]. Shih W-H, Shih WY, Kim S-I, Liu J, Aksay IA. Scaling behavior of the elastic properties of colloidal gels. *Physical Review A*. 1990; 42:4772–4779.
- [47]. Piau J, Dorget M, Palierne J. Shear elasticity and yield stress of silica-silicone physical gels: Fractal approach. *Journal of Rheology*. 1999; 43:305–314.
- [48]. Wu H, Morbidelli M. A model relating structure of colloidal gels to their elastic properties. *Langmuir*. 2001; 17:1030–1036.
- [49]. Xi Y, Chen J, Xu Y, Chu F, Liu C. Yield Stress of Fractal Aggregates. *Fractals*. 2015; 23:1550028.

- [50]. Urieu N, Ladyzhinsky IY. Fractal models in rheology of colloidal gels. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 1996; 108:1–11.
- [51]. Studart AR, Amstad E, Gauckler LJ. Yielding of weakly attractive nanoparticle networks. *Soft Matter*. 2011; 7:6408–6412.
- [52]. Shih WY, Shih WH, Aksay IA. Elastic and yield behavior of strongly flocculated colloids. *Journal of the American Ceramic Society*. 1999; 82:616–624.
- [53]. Wessel R, Ball R. Fractal aggregates and gels in shear flow. *Physical Review A*. 1992; 46:R3008–R3011.
- [54]. De Gennes, P-G. *Scaling concepts in polymer physics*. Cornell University Press; Ithaca: 1979.
- [55]. Saint-Michel F, Pignon F, Magnin A. Fractal behavior and scaling law of hydrophobic silica in polyol. *Journal of colloid and interface science*. 2003; 267:314–319. [PubMed: 14583206]
- [56]. Loiseau A, Tassin J-F. Model nanocomposites based on laponite and poly (ethylene oxide): preparation and rheology. *Macromolecules*. 2006; 39:9185–9191.
- [57]. Gareche M, Allal A, Zeraibi N, Roby F, Azril N, Saoudi L. Relationship between the fractal structure with the shear complex modulus of montmorillonite suspensions. *Applied Clay Science*. 2016; 123:11–17.
- [58]. Zaccone A, Wu H, Del Gado E. Elasticity of arrested short-ranged attractive colloids: Homogeneous and heterogeneous glasses. *Physical review letters*. 2009; 103:208301. [PubMed: 20366015]
- [59]. Wyss HM, Tervoort E, Meier LP, Müller M, Gauckler LJ. Relation between microstructure and mechanical behavior of concentrated silica gels. *Journal of colloid and interface science*. 2004; 273:455–462. [PubMed: 15082380]
- [60]. Pignon F, Magnin A, Piau J-M, Cabane B, Lindner P, Diat O. Yield stress thixotropic clay suspension: Investigations of structure by light, neutron, and x-ray scattering. *Physical Review E*. 1997; 56:3281–3289.
- [61]. Stöber W, Fink A, Bohn E. Controlled growth of monodisperse silica spheres in the micron size range. *Journal of Colloid and Interface Science*. 1968; 26:62–69.
- [62]. Nozawa K, Gailhanou H, Raison L, Panizza P, Ushiki H, Sellier E, Delville J, Delville M. Smart control of monodisperse Stöber silica particles: effect of reactant addition rate on growth process. *Langmuir*. 2005; 21:1516–1523. [PubMed: 15697302]
- [63]. Fuji M, Fujimori H, Takei T, Watanabe T, Chikazawa M. Wettability of glass-bead surface modified by trimethylchlorosilane. *The Journal of Physical Chemistry B*. 1998; 102:10498–10504.
- [64]. Brummer, R. *Rheology essentials of cosmetic and food emulsions*. Springer Science & Business Media; 2006.
- [65]. Maurath J, Bitsch B, Schwegler Y, Willenbacher N. Influence of particle shape on the rheological behavior of three-phase non-brownian suspensions. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2016; 497:316–326.
- [66]. Crocker JC, Grier DG. Methods of digital video microscopy for colloidal studies. *Journal of Colloid and Interface Science*. 1996; 179:298–310.
- [67]. Weeks ER, Crocker JC, Levitt AC, Schofield A, Weitz DA. Three-dimensional direct imaging of structural relaxation near the colloidal glass transition. *Science*. 2000; 287:627–631. [PubMed: 10649991]
- [68]. Crocker, JC., Weeks, ER. Particle tracking using IDL. Retrieved from <http://www.physics.emory.edu/faculty/weeks/idl/>
- [69]. Wenzl J, Seto R, Roth M, Butt H-J, Auernhammer GK. Measurement of rotation of individual spherical particles in cohesive granulates. *Granular Matter*. 2012; 15:391–400.
- [70]. Scheel M, Seemann R, Brinkmann M, Di Michiel M, Sheppard A, Breidenbach B, Herminghaus S. Morphological clues to wet granular pile stability. *Nature Materials*. 2008; 7:189–193. [PubMed: 18264104]
- [71]. Yang J, Velankar SS. Preparation and yielding behavior of pendular network suspensions. *Journal of Rheology*. 2017; 61:217–228.

- [72]. Torres FE, Russel WB, Schowalter WR. Floc Structure and Growth Kinetics for Rapid Shear Coagulation of Polystyrene Colloids. *Journal of colloid and interface science*. 1991; 142:554–574.
- [73]. Lin M, Klein R, Lindsay H, Weitz D, Ball R, Meakin P. The structure of fractal colloidal aggregates of finite extent. *Journal of colloid and interface science*. 1990; 137:263–280.
- [74]. Allain C, Cloitre M, Parisse F. Settling by cluster deposition in aggregating colloidal suspensions. *Journal of colloid and interface science*. 1996; 178:411–416.
- [75]. Allain C, Cloitre M, Wafra M. Aggregation and sedimentation in colloidal suspensions. *Physical review letters*. 1995; 74:1478–1481. [PubMed: 10059030]
- [76]. Peltomäki M, Hellén E, Alava M. No self-similar aggregates with sedimentation. *Journal of Statistical Mechanics: Theory and Experiment*. 2004 P09002.
- [77]. Lach-Hab M, González AE, Blaisten-Barojas E. Concentration dependence of structural and dynamical quantities in colloidal aggregation: computer simulations. *Physical Review E*. 1996; 54:5456–5462.
- [78]. Au P-I, Liu J, Leong Y-K. Yield stress and microstructure of washed oxide suspensions at the isoelectric point: experimental and model fractal structure. *Rheologica Acta*. 2016; 55:847–856.
- [79]. Ikeda S, Foegeding EA, Hagiwara T. Rheological study on the fractal nature of the protein gel structure. *Langmuir*. 1999; 15:8584–8589.
- [80]. Larson, RG. *The Structure and Rheology of Complex Fluids*. Oxford University Press; Oxford: 1999.
- [81]. Dullaert K, Mewis J. Thixotropy: Build-up and breakdown curves during flow. *Journal of Rheology*. 2005; 49:1213–1230.
- [82]. Varadan P, Solomon MJ. Direct visualization of long-range heterogeneous structure in dense colloidal gels. *Langmuir*. 2003; 19:509–512.
- [83]. Dinsmore A, Weitz D. Direct imaging of three-dimensional structure and topology of colloidal gels. *Journal of Physics: Condensed Matter*. 2002; 14:7581–7597.
- [84]. Colombo J, Del Gado E. Stress localization, stiffening, and yielding in a model colloidal gel. *Journal of Rheology*. 2014; 58:1089–1116.

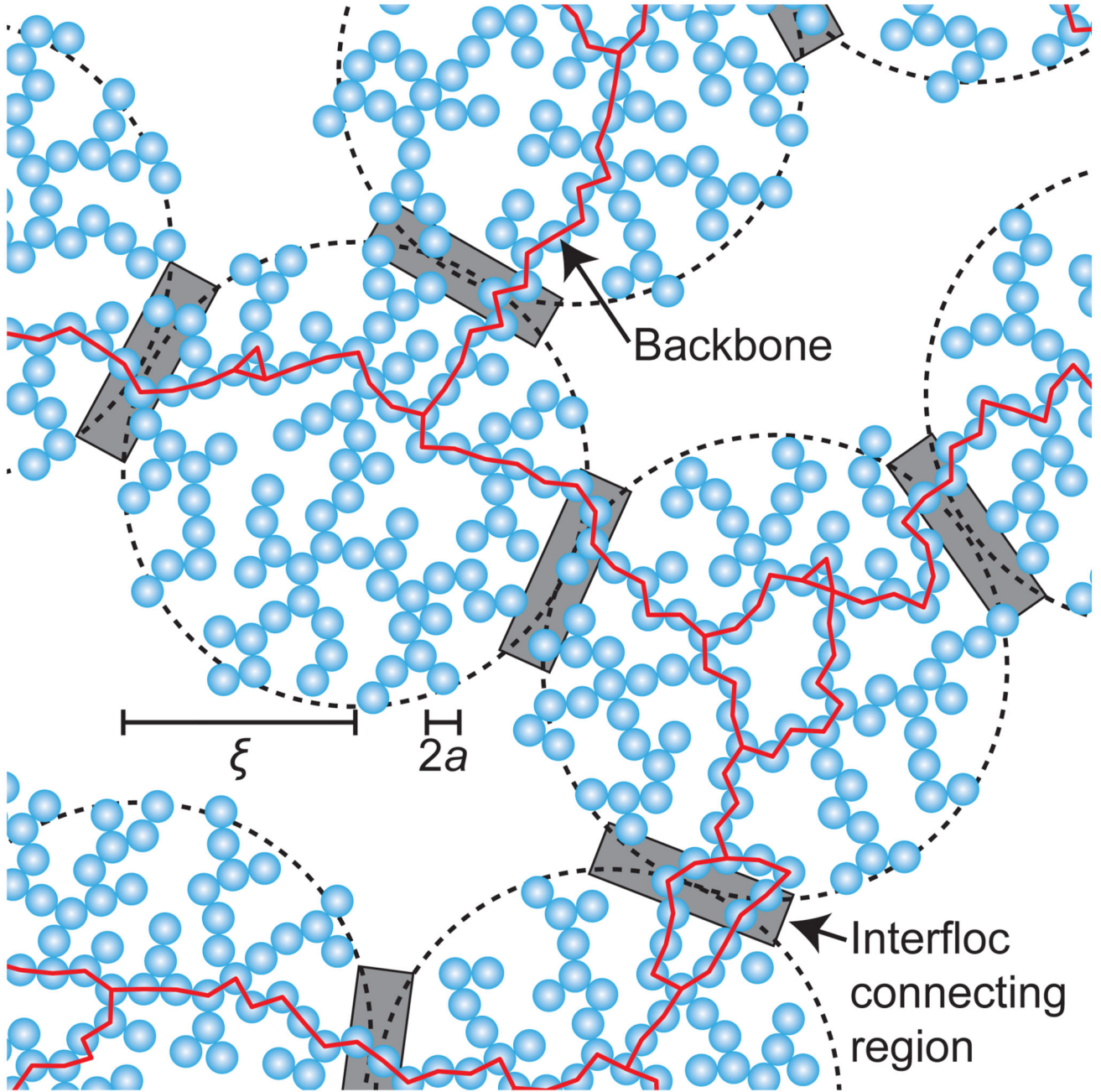


Figure 1.

Schematic drawing of a percolated network consisting of interconnected clusters (flocs) of particles having radius a . The large dashed-line circles denote flocs with correlation length ξ and interfloc connecting regions are highlighted by the grey shaded areas. A backbone transmitting forces in the particle network is indicated by the thick red lines. While most interfloc links are part of the backbone, only a minor amount of the total particles inside of a floc contributes to the backbone.

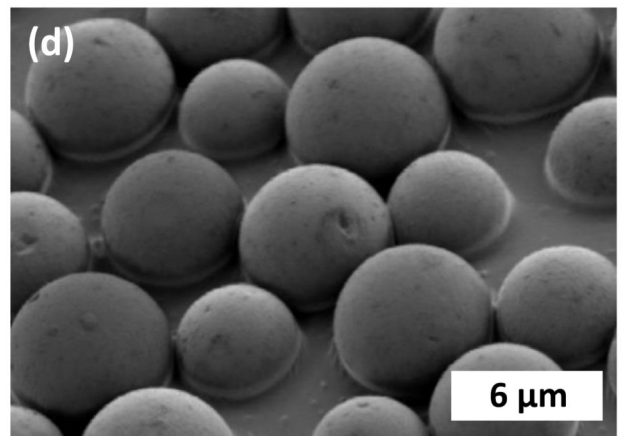
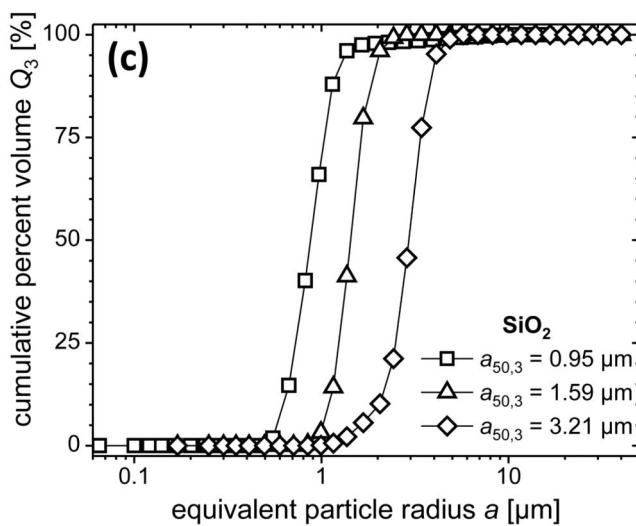
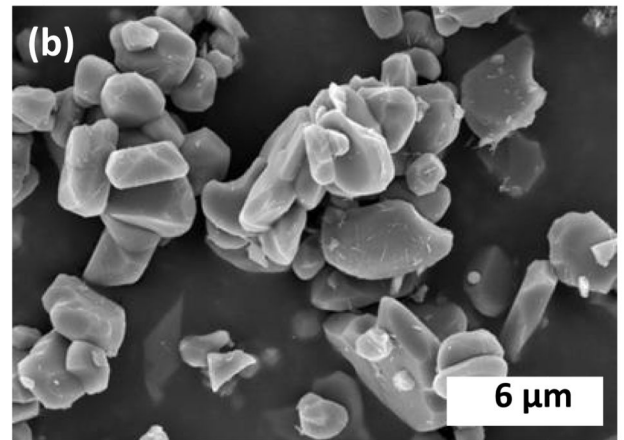
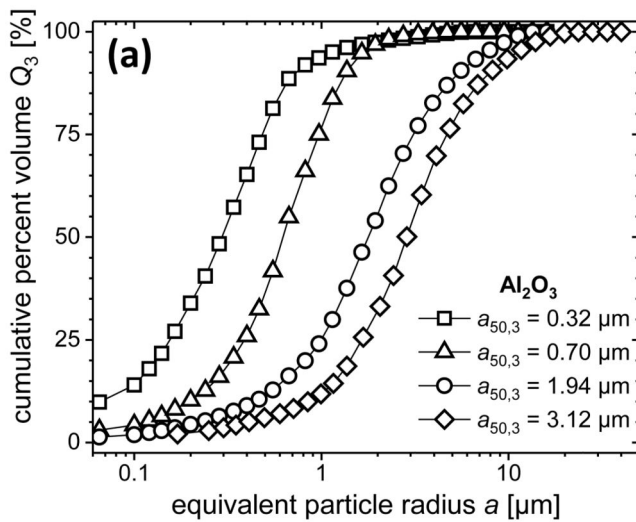


Figure 2. Particle size distributions and scanning electron microscopy (SEM) images of both particle types used in this study. (a) Particle size distribution of the different Al_2O_3 particles. (b) SEM image of Al_2O_3 with mean radius $a_{50,3} = 3.12 \mu\text{m}$. (c) Particle size distribution of the different silica particles. (d) SEM image of silica with $a_{50,3} = 3.21 \mu\text{m}$.

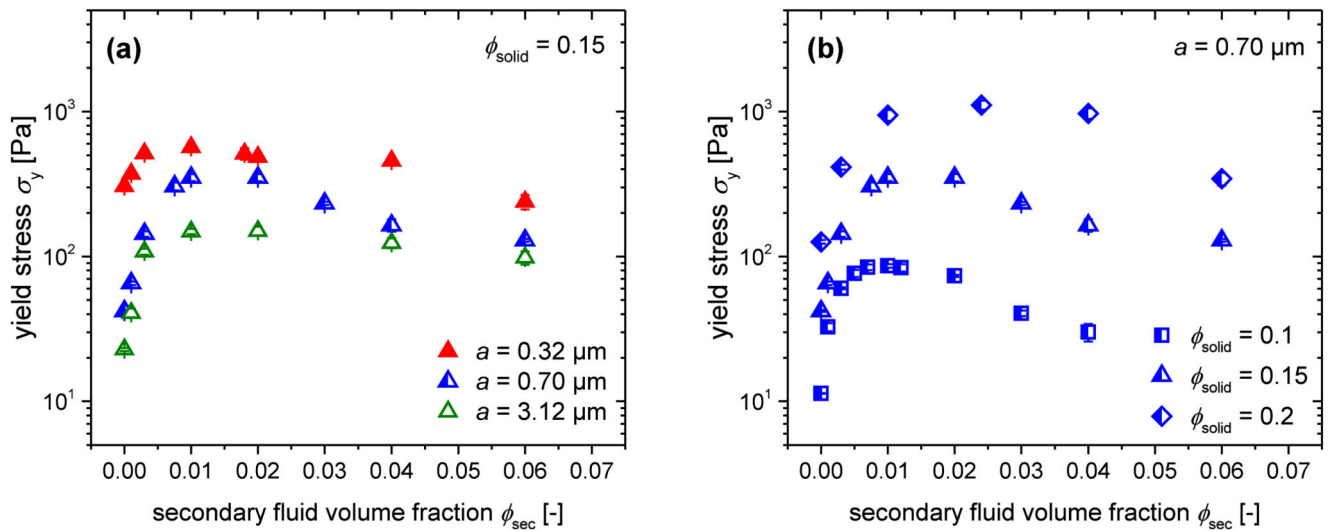


Figure 3.

Yield stress σ_y as function of second fluid volume fraction ϕ_{sec} for Al_2O_3 based capillary suspensions at (a) three different particle sizes a with constant solid volume fraction $\phi_{solid} = 0.15$ and (b) three different ϕ_{solid} with constant particle size $a = 0.70 \mu\text{m}$.

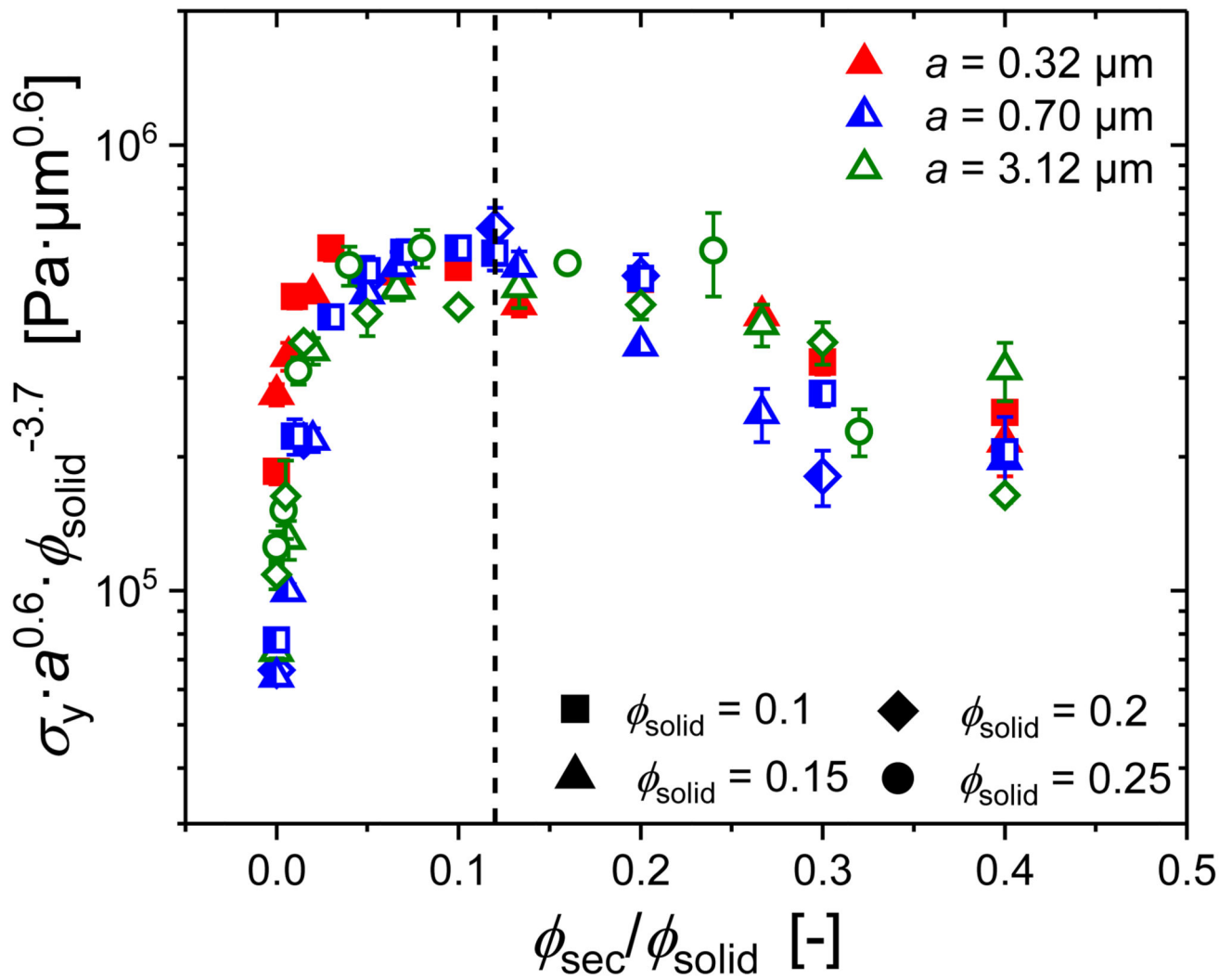


Figure 4. Yield stress master curve for Al₂O₃ based capillary suspensions. Different symbol shapes denote each solid volume fraction and symbol fillings and color denote each particle size. The dashed line designates the position of the yield stress maximum at $\phi_{\text{sec}}/\phi_{\text{solid}} = 0.12$.

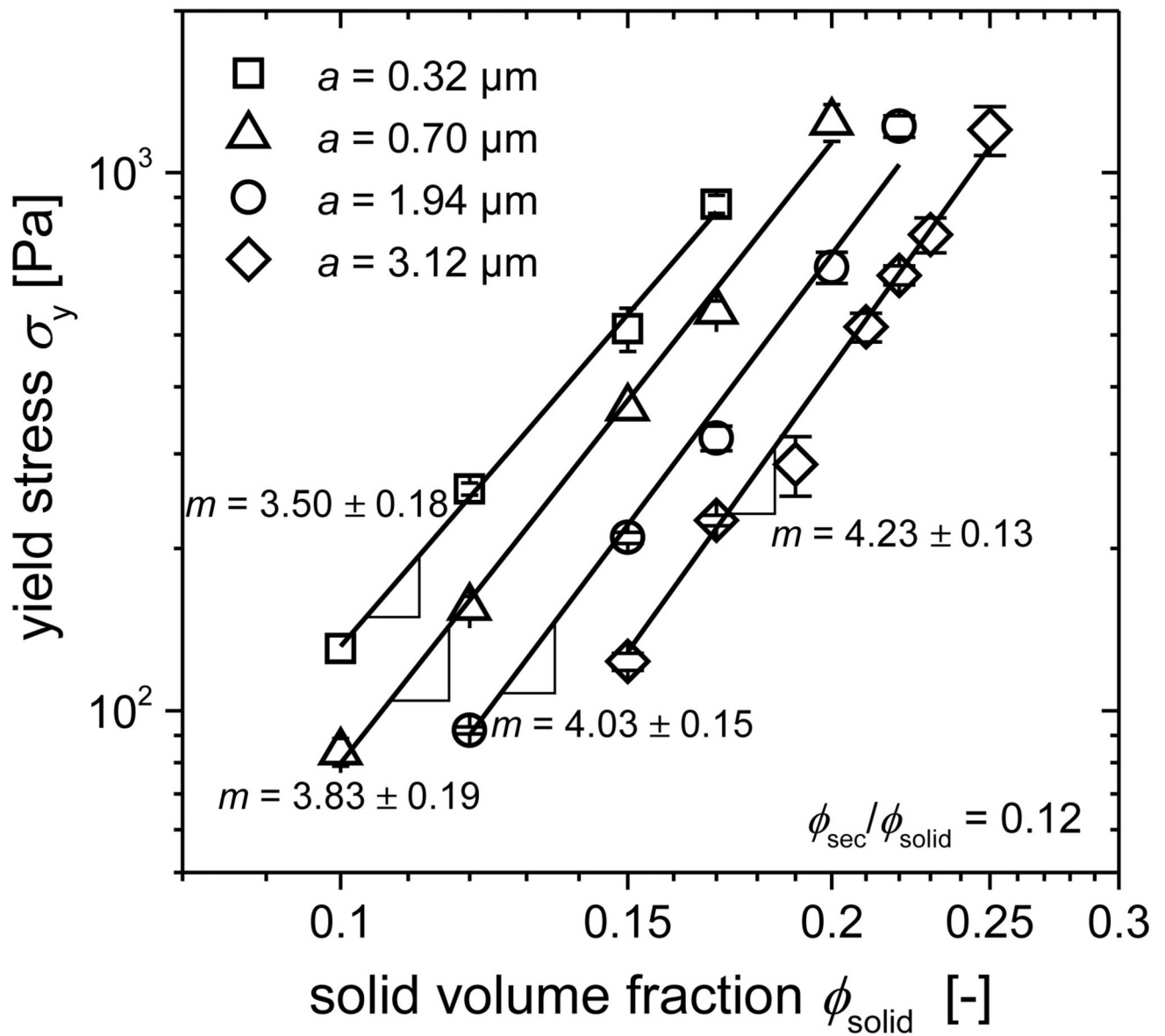


Figure 5. Yield stress σ_y of Al_2O_3 based capillary suspensions as function of the solid volume fraction ϕ_{solid} at a fixed ratio of $\phi_{\text{sec}} / \phi_{\text{solid}} = 0.12$ for the four different mean particle radii.

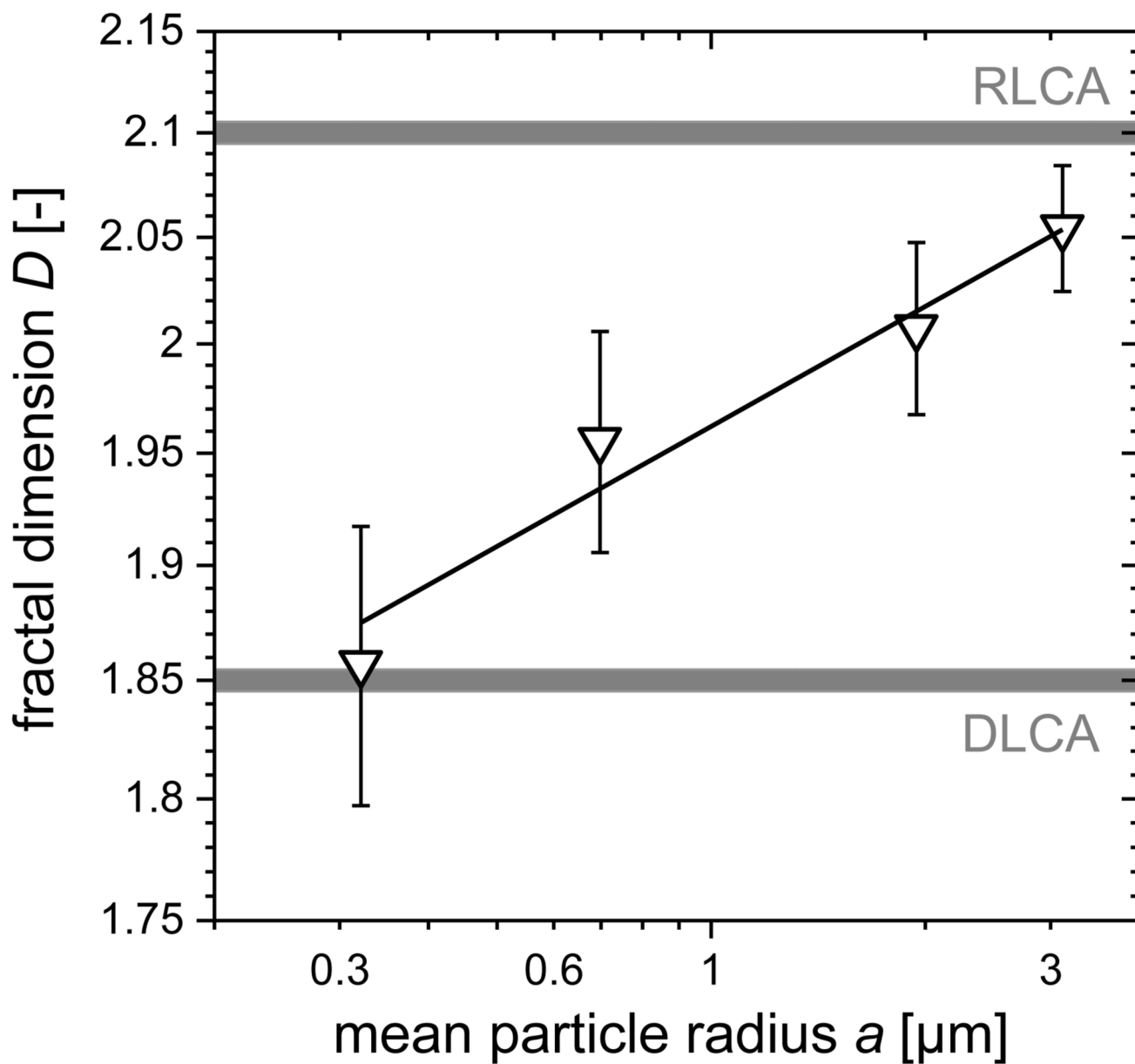


Figure 6. Fractal dimension D as function of the particle size calculated with the Piau yield stress scaling model. The grey horizontal lines denote expected fractal dimension values for DLCA and RLCA aggregation.

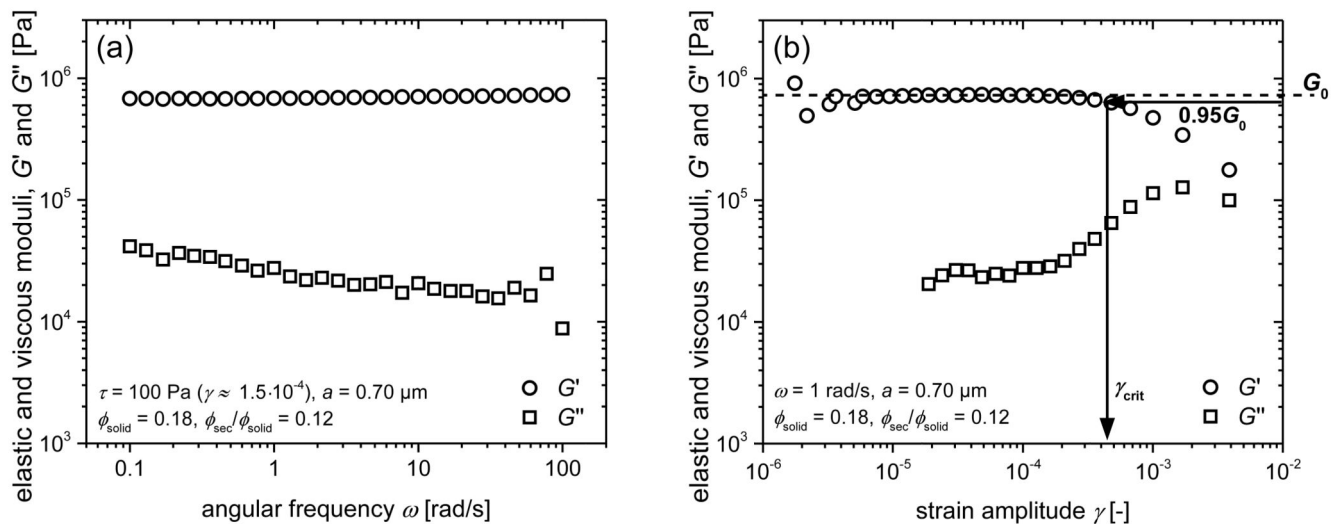
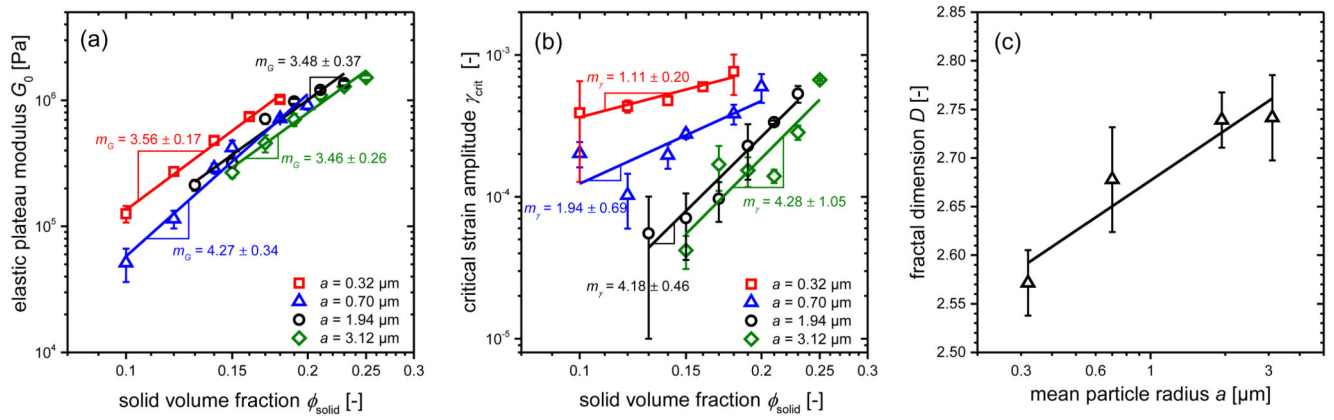


Figure 7.

Elastic and viscous shear moduli G' and G'' as a function of (a) angular frequency ω at a stress (and strain) amplitude inside the linear viscoelastic region and (b) strain amplitude at a frequency $\omega = 1$ rad/s. The elastic modulus G' is shown as open circles and the viscous modulus G'' as open squares. The dotted line in (b) denotes the elastic plateau modulus G_0 while the arrows illustrate the determination of the critical strain amplitude γ_{crit} . Example data of an Al_2O_3 -based capillary suspension with $a = 0.70$ μm , $\phi_{\text{solid}} = 0.18$ and $\phi_{\text{sec}}/\phi_{\text{solid}} = 0.12$.

**Figure 8.**

(a) Elastic plateau modulus G_0 and (b) critical strain amplitude γ_{crit} of Al₂O₃-based capillary suspensions as function of the solid volume fraction ϕ_{solid} at a fixed ratio of $\phi_{\text{sec}}/\phi_{\text{solid}} = 0.12$. (c) Fractal dimensions D as function of the mean particle size calculated from the data shown in (a) and (b) using the model of Wu and Morbidelli.

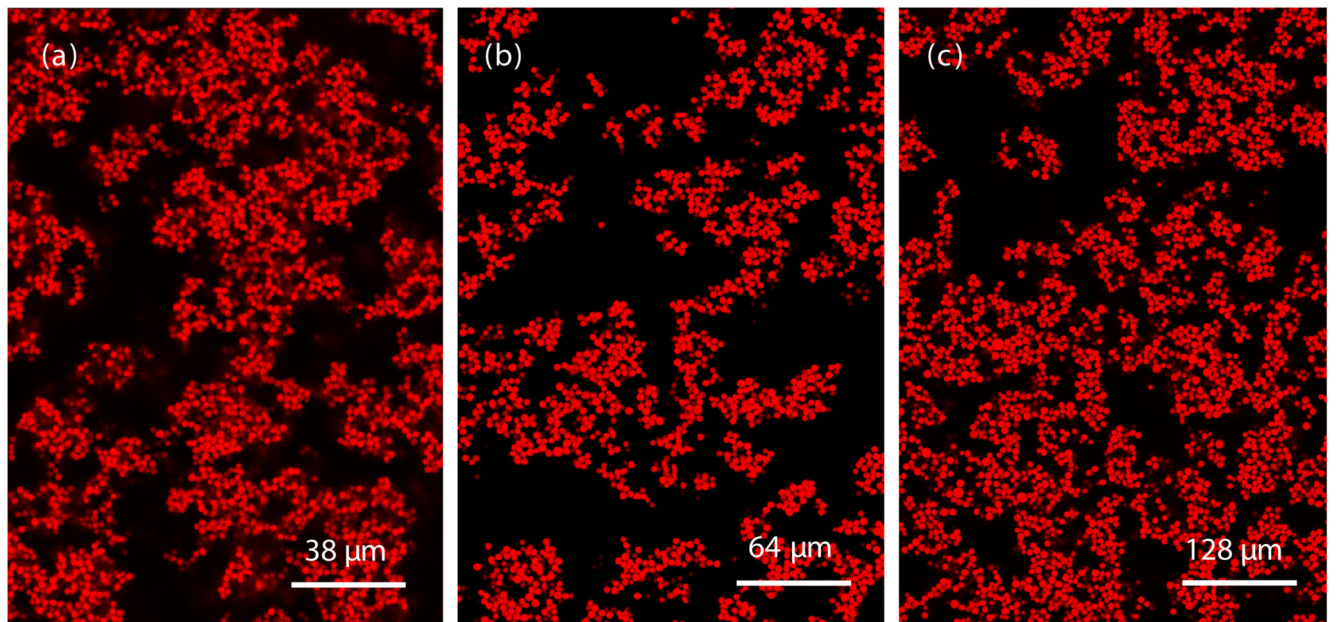


Figure 9.

Confocal 2D images of capillary suspensions based on silica particles ($\phi_{\text{solid}} = 0.26 \pm 0.06$) with three different mean particle radii (a) $a = 0.95 \mu\text{m}$, (b) $a = 1.59 \mu\text{m}$ and (c) $a = 3.21 \mu\text{m}$, all at $\phi_{\text{sec}}/\phi_{\text{solid}} = 0.12$. The silica particles are shown in red. The undyed bulk liquid and secondary liquid are not shown. Image sizes and scale bars have been adjusted so that the scale bar equals $40 \cdot a$ in each image.

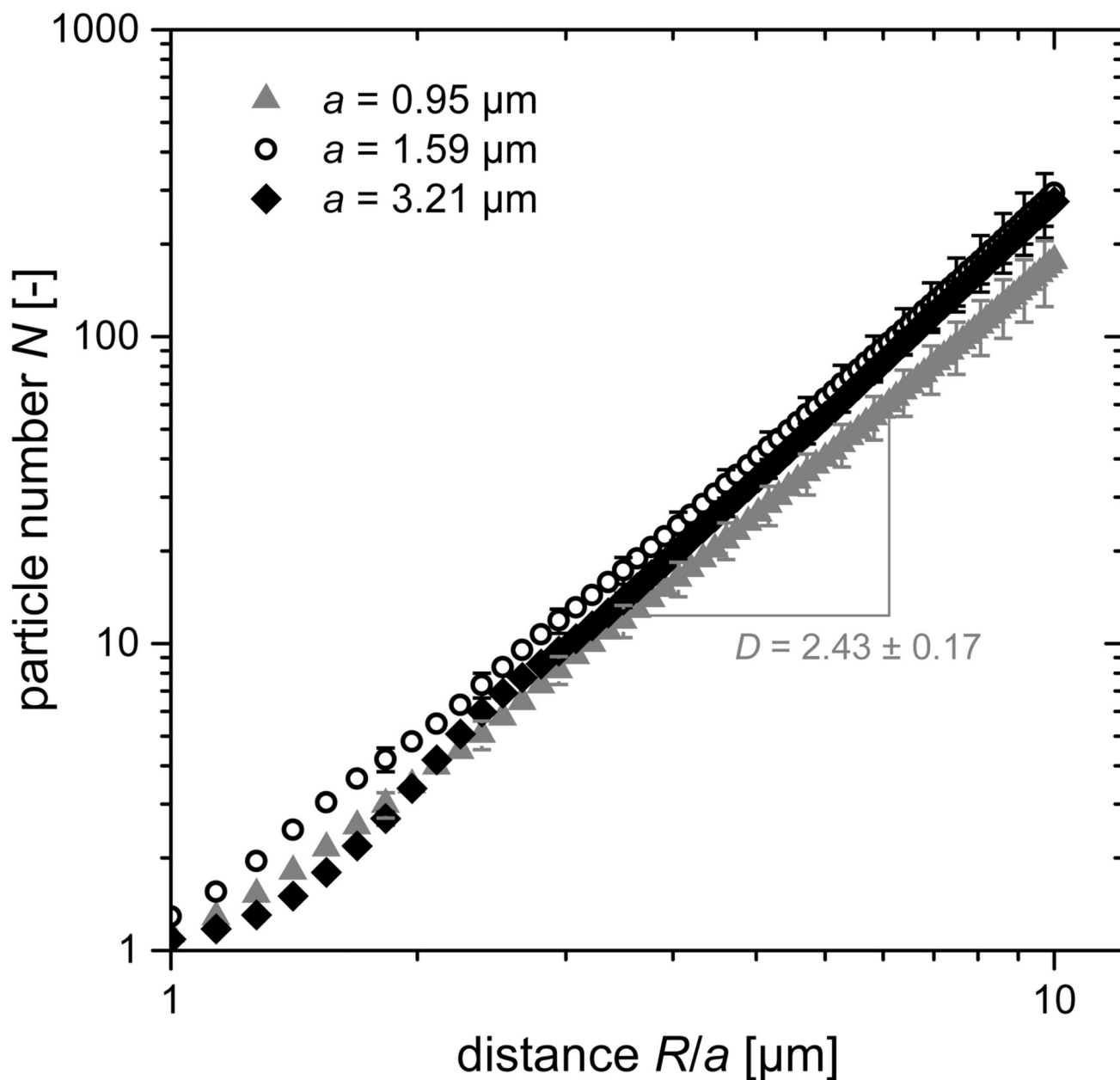


Figure 10.

Particle number function $N(R/a)$ for silica based capillary suspensions with $a = 0.95 \mu\text{m}$ (grey triangles), $a = 1.59 \mu\text{m}$ (black open circles) and $a = 3.21 \mu\text{m}$ (black filled diamonds), all with $\phi_{\text{solid}} = 0.26 \pm 0.06$ and $\phi_{\text{sec}}/\phi_{\text{solid}} = 0.12 \pm 0.03$ as determined from 3D confocal image data. Error bars are only shown at every fourth data point for the sake of clarity. The exponent of the $N(R/a)$ function equals the fractal dimension D as exemplified in the figure for the particles with $a = 0.95 \mu\text{m}$.

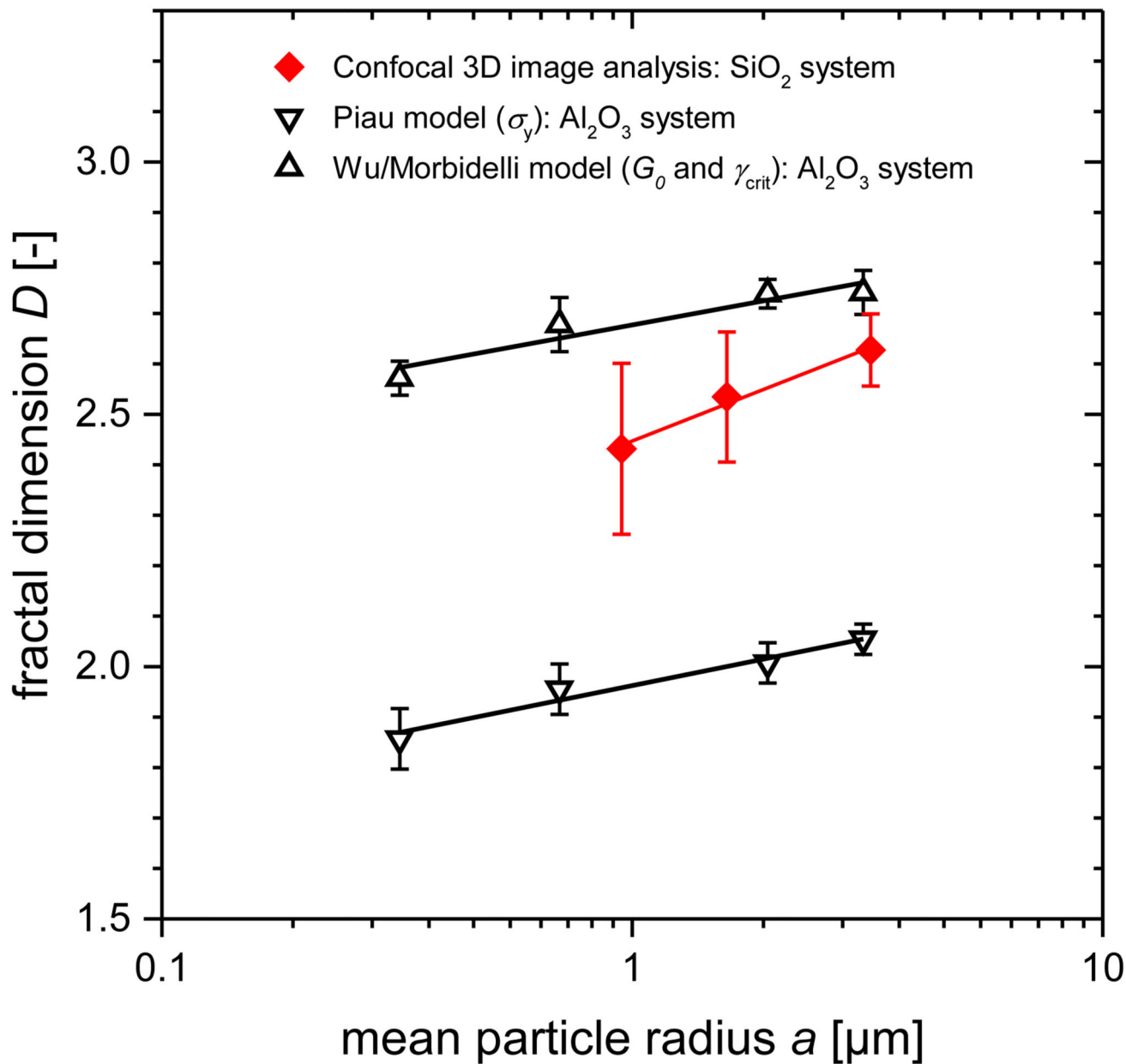


Figure 11.

Apparent fractal dimensions D as function of particle size a determined from confocal image analysis of silica based capillary suspensions (red diamonds). For comparison, the results from the rheological scaling models presented in section 4.2 are reproduced in this diagram, where the backbone dimension was calculated using the model of Piau et al and the aggregate dimension with the model of Wu and Morbidelli.